

Total Syntheses of Nannocystins A and A0, Two Elongation Factor 1 **Inhibitors**

Jun Huang and Zhang Wang*

Department of Chemistry, University at Albany, State University of New York, Albany, New York 12222, United States

Supporting Information

ABSTRACT: Asymmetric total syntheses of nannocystins A and A0 were achieved in a convergent route starting from simple materials. Nannocystin family natural products bear potent anticancer activity as elongation factor 1 inhibitors. In this synthesis, the challenging tertiary amide bond was constructed by peptide coupling between an acyl chloride and a secondary amine. A late-stage ring-closing metathesis reaction successfully rendered the macrocycle. This efficient synthetic strategy should be applicable to other nannocystins and analogues and therefore should benefit future structure-activity relationship studies.

Tatural products serve as a main source of therapeutics because of their unique structures and structural diversity. About half of the current small-molecule drugs on the market are natural products or their synthetic analogues. When new natural products with unprecedented structural features are isolated, scientists often find new mechanisms of action for these natural products in biological systems, which could lead to new drug development, as exemplified by research on calicheamicin and rapamycin.² In June 2015, in searching for new bioactive natural products, Hoffmann et al.3 reported a family of cyclodepsipeptides called nannocystins (Figure 1) with potent antiproliferative activities against a number of cancer cell lines at nanomolar concentrations. For example, the IC₅₀'s of nannocystin A (1) against HCT116, PC3, and HL60 are 1.2, 1.0, and 12 nM, respectively. Almost simultaneously,

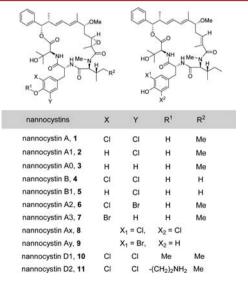


Figure 1. Natural nannocystins (1-9) and their synthetic analogues (10 and 11).

Krastel et al.⁴ also discovered the nannocystins and conducted a more comprehensive investigation. They found that 1 showed antiproliferative properties against 472 cancer cell lines in the nanomolar concentration range. Moreover, a series of experimental and computational studies by Krastel et al. strongly suggested that the target protein of nannocystins is elongation factor $1-\alpha$ (EF- 1α). Elongation factors secure accuracy in the translation process and are important in protein synthesis.⁵ Cancer cells tend to overexpress elongation factors to expedite protein production. Therefore, compounds that target elongation factors, such as nannocystins, may serve as lead candidates for anticancer therapy. Attracted by their potent biological activity and urged by subsequent structureactivity relationship studies, we embarked on the total synthesis of nannocystins. Here we report the first asymmetric total syntheses of nannocystins A (1) and A0 (3).

We sought a convergent synthesis of nannocystins. Our retrosynthetic analysis is shown in Scheme 1. We planned to take advantage of the ring-closing metathesis (RCM) reaction of 12 to assemble the macrocycle. The RCM strategy for macrocycle synthesis is a robust method, as documented in the literature.⁸ Disconnection of the tertiary amide bond of 12 would lead to peptide fragment 13 and polyketide motif 14 with approximately equal complexity. Compound 13 can be derived from ester 15 through a peptide coupling reaction, and ester 15 should be easily prepared from homoallylic alcohol 16. On the other hand, carboxylic acid 14 could be made via an asymmetric epoxidation of alcohol 17 followed by oxidation. Allylic alcohol 17 could be made via a regioselective crossmetathesis reaction of the simple compound 18, the asymmetric allylation product of (*E*)-3-bromomethacrolein.

The synthesis of acid 14 started from the known compound (E)-3-bromomethacrolein (19) (Scheme 2). Enantioselective allylation of 19 using Antilla's method provided alcohol 20 in

Received: August 6, 2016 Published: September 6, 2016 Organic Letters Letter

Scheme 1. Retrosynthetic Analysis

Scheme 2. Synthesis of Acid 14

97% yield with 98% ee. 10 The optical purity of **20** was determined from its UV-active derivative **17** (vide infra). The free hydroxyl group in **20** was methylated by treatment with sodium hydride and iodomethane in 95% yield. Methyl ether **18** underwent cross-metathesis with the monosubstituted olefin regio- and stereoselectively using Grubbs' second-generation catalyst and neat methacrolein as the solvent, 11 furnishing enal **21** in 61% yield. A prolonged reaction time (36 h) and excess methacrolein was necessary to obtain a good yield of **21** in this step. Luche reduction of enal **21** in ethanol smoothly rendered allylic alcohol **22** in 72% yield. 12 The next goal was to introduce the diene functionality via cross-coupling reactions. We found the more active palladium catalyst Pd(Pt-Bu₃)₂ to be superior to Pd(PPh₃)₄ in the Stille coupling reaction with tri-nbutylvinylstannane, furnishing diene **17** in 82% yield. 13

Sharpless asymmetric epoxidation of allylic alcohol 17 gave the desired epoxide 23 in 95% yield with amplified optical purity (>99% ee). Finally, direct oxidation of alcohol 23 to acid 14 using iodosobenzene diacetate and a catalytic amount of 2-azaadamantane-*N*-oxyl (AZADO) successfully furnished acid 14 in 55% yield. Sharpless asymmetric epoxidation of allylic alcohol 23 to acid 14 in 55% yield.

The assembly of peptide motif 13 started with the synthesis of 15 via Mitsunobu reaction between the known compounds 16 and 24 (Scheme 3). Deprotection of *tert*-butyl

Scheme 3. Synthesis of Peptide Motif 13

carbamate 15 furnished ammonium salt 25. *N*-Fmoc-protected *N*-methyl-L-isoleucine (26) was successfully coupled with D-tyrosine benzyl ester, affording dipeptide 27 in 95% yield. After hydrogenolysis of the benzyl group, acid 28 was connected with 25 using HBTU, giving tripeptide 29 in 60% yield. The free secondary amine 13 was produced by treatment of 29 with diethylamine in dry acetonitrile.

With fragments 13 and 14 in hand, we turned our attention to the coupling of the two termini of 13 and 14 to finish the total synthesis (Scheme 4). The carbonyl group in acid 14 is quite hindered because of the tetrasubstituted α -carbon. At the same time, isoleucine is a hindered amino acid in peptide

Scheme 4. Total Synthesis of Nannocystin A0

Organic Letters Letter

coupling, especially when the nitrogen atom is alkylated.¹⁸ Common peptide coupling reagents such as HATU, HBTU, EDCI, etc., failed to generate the coupled product 12. Therefore, we focused on stronger acyl donors generated from 14. Happily, we were able to make the corresponding acyl chloride 31 using Ghosez reagent 30.19 Crude 31 in a dichloromethane solution was added dropwise to 13 in dichloromethane at -20 °C, and the desired product 12 was obtained in 80% yield.²⁰ It should be noted that compounds 29, 13, and 14 had to be purified by preparative TLC to remove small amounts of impurities. In addition, compounds 14 and 31 are not very stable, probably because of the acid-sensitive diene moiety, and had to be used immediately after preparation. Finally, the target natural product nannocystin A0 (3) was synthesized successfully via ring-closing metathesis of 12 using the second-generation Hoveyda-Grubbs catalyst in 79% yield as a 4.6:1 ratio of isomers (3 is the major isomer).²¹ The functional group compatibility of this metathesis macrocyclization step is quite remarkable. Pure 3 was obtained after separation by reversed-phase HPLC. The NMR spectra of synthetic 3 were identical to those of natural nannocystin A0 reported by Krastel et al.4

The synthetic sequence for nannocystin A0 could be generalized to the synthesis of other nannocystins. Starting from 3,5-dichloro-D-tyrosine benzyl ester,²² we prepared nannocystin A (1) in the same manner (Scheme 5; also see the Supporting Information). The NMR spectra of synthetic 1 also matched those of natural nannocystin A reported by Krastel et al.⁴

Scheme 5. Highlights of the Total Synthesis of Nannocystin A (1)

In summary, convergent total syntheses of nannocystins A and A0 were achieved in nine steps (longest linear sequence) from a simple starting material, (*E*)-3-bromomethacrolein. Noteworthy steps include the peptide coupling between a reactive acyl chloride and a hindered *N*-methyl-L-isoleucine moiety and the ring-closing metathesis macrocyclization. Our synthetic route will benefit future structure—activity relationship studies of nannocystins.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.orglett.6b02352.

Detailed experimental procedures and characterizations of new compounds (PDF) NMR data (ZIP)

AUTHOR INFORMATION

Corresponding Author

*E-mail: zwang9@albany.edu.

Notes

The authors declare the following competing financial interest(s): A patent application was filed by University at Albany, State University of New York.

ACKNOWLEDGMENTS

We are grateful to University at Albany, State University of New York, for financial support. We thank Prof. Alexander Shekhtman (SUNY-Albany), Dr. Xiaochuan Cai (Memorial Sloan-Kettering Cancer Center), and Dr. Rong Long (Columbia University) for NMR assistance. We also thank Dr. Philipp Krastel (Novartis Institutes for BioMedical Research) for providing the NMR spectra of natural nannocystins A and A0. Dr. Ying Wu is acknowledged for helping prepare this article. This work is dedicated to Professor Samuel J. Danishefsky (Columbia University and Memorial Sloan-Kettering Cancer Center) on the occasion of his 80th birthday.

REFERENCES

- (1) For selected reviews of natural products and drugs, see: (a) Nicolaou, K. C.; Montagnon, T. Molecules That Changed the World; Wiley-VCH: Weinheim, Germany, 2008. (b) Dias, D. A.; Urban, S.; Roessner, U. Metabolites 2012, 2, 303. (c) Szychowski, J.; Truchon, J.-F.; Bennani, Y. L. J. Med. Chem. 2014, 57, 9292. (d) Gogineni, V.; Schinazi, R. F.; Hamann, M. T. Chem. Rev. 2015, 115, 9655. (e) Mishra, B. B.; Tiwari, V. K. Eur. J. Med. Chem. 2011, 46, 4769. (f) Newman, D. J.; Cragg, G. M. J. Nat. Prod. 2016, 79, 629.
- (2) For reviews of calicheamicins, see: (a) Lee, M. D.; Ellestad, G. A.; Borders, D. B. Acc. Chem. Res. 1991, 24, 235. (b) Ellestad, G. A. Chirality 2011, 23, 660. (c) Bross, P. F.; Beitz, J.; Chen, G.; Chen, X. H.; Duffy, E.; Kieffer, L.; Roy, S.; Sridhara, R.; Rahman, A.; Williams, G.; Pazdur, R. Clin. Cancer Res. 2001, 7, 1490. For reviews of rapamycin, see: (d) Heitman, J.; Movva, N. R.; Hall, M. N. Science 1991, 253, 905. (e) Foster, K. G.; Fingar, D. C. J. Biol. Chem. 2010, 285, 14071. (f) Easton, J. B.; Houghton, P. J. Oncogene 2006, 25, 6436.
- (3) Hoffmann, H.; Kogler, H.; Heyse, W.; Matter, H.; Caspers, M.; Schummer, D.; Klemke-Jahn, C.; Bauer, A.; Penarier, G.; Debussche, L.; Brönstrup, M. *Angew. Chem., Int. Ed.* **2015**, *54*, 10145.
- (4) Krastel, P.; Roggo, S.; Schirle, M.; Ross, N. T.; Perruccio, F.; Aspesi, P., Jr.; Aust, T.; Buntin, K.; Estoppey, D.; Liechty, B.; Mapa, F.; Memmert, K.; Miller, H.; Pan, X.; Riedl, R.; Thibaut, C.; Thomas, J.; Wagner, T.; Weber, E.; Xie, X.; Schmitt, E. K.; Hoepfner, D. Angew. Chem., Int. Ed. 2015, 54, 10149.
- (5) For reviews of EF-1α, see: (a) Sasikumar, A. N.; Perez, W. B.; Kinzy, T. G. WIREs RNA **2012**, 3, 543. (b) Dever, T. E.; Green, R. Cold Spring Harbor Perspect. Biol. **2012**, 4, a013706.
- (6) (a) Pecorari, L.; Marin, O.; Silvestri, C.; Candini, O.; Rossi, E.; Guerzoni, C.; Cattelani, S.; Mariani, S. A.; Corradini, F.; Ferrari-Amorotti, G.; Cortesi, L.; Bussolari, R.; Raschellà, G.; Federico, M. R.; Calabretta, B. *Mol. Cancer* 2009, 8, 58. (b) Lamberti, A.; Caraglia, M.; Longo, O.; Marra, M.; Abbruzzese, A.; Arcari, P. *Amino Acids* 2004, 26, 443. (c) Dua, K.; Williams, T. M.; Beretta, L. *Proteomics* 2001, 1, 1191. (d) Lee, J. M. *Reprod. Biol. Endocrinol.* 2003, 1, 69.
- (7) This work was presented at the 12th SINO-US Chemistry Professors Conference in Guangzhou, China, on June 25, 2016.
- (8) For reviews, see: (a) Gradillas, A.; Pérez-Castells, J. Angew. Chem., Int. Ed. 2006, 45, 6086. (b) Nicolaou, K. C.; Bulger, P. G.; Sarlah, D. Angew. Chem., Int. Ed. 2005, 44, 4490. (c) Prunet, J. Eur. J. Org. Chem. 2011, 2011, 3634. (d) Hoveyda, A. H.; Zhugralin, A. R. Nature 2007, 450, 243. (e) Fürstner, A. Chem. Commun. 2011, 47, 6505.
- (9) Wang, Y.; Dai, W.-M. Eur. J. Org. Chem. 2014, 2014, 323.

Organic Letters Letter

- (10) Jain, P.; Antilla, J. C. J. Am. Chem. Soc. 2010, 132, 11884.
- (11) (a) Xu, S.; Arimoto, H.; Uemura, D. Angew. Chem., Int. Ed. **2007**, 46, 5746. (b) Xu, J.; Caro-Diaz, E. J. E.; Trzoss, L.; Theodorakis, E. A. J. Am. Chem. Soc. **2012**, 134, 5072.
- (12) Luche, J.-L. J. Am. Chem. Soc. 1978, 100, 2226.
- (13) (a) Littke, A. F.; Schwarz, L.; Fu, G. C. J. Am. Chem. Soc. 2002, 124, 6343. (b) Han, X.; Stoltz, B. M.; Corey, E. J. J. Am. Chem. Soc. 1999, 121, 7600.
- (14) Gao, Y.; Hanson, R. M.; Klunder, J. M.; Ko, S. Y.; Masamune, H.; Sharpless, K. B. *J. Am. Chem. Soc.* **1987**, *109*, 5765.
- (15) (a) Shibuya, M.; Tomizawa, M.; Suzuki, I.; Iwabuchi, Y. J. Am. Chem. Soc. 2006, 128, 8412. (b) Sun, Y.; Chen, P.; Zhang, D.; Baunach, M.; Hertweck, C.; Li, A. Angew. Chem., Int. Ed. 2014, 53, 9012. (c) Kuranaga, T.; Sesoko, Y.; Sakata, K.; Maeda, N.; Hayata, A.; Inoue, M. J. Am. Chem. Soc. 2013, 135, 5467.
- (16) Dettwiler, J. E.; Bélec, L.; Lubell, W. D. Can. J. Chem. 2005, 83, 793.
- (17) For selected methods for the asymmetric synthesis of compound 16, see ref 9 and: (a) Brown, H. C.; Bhat, K. S. J. Am. Chem. Soc. 1986, 108, 5919. (b) Roush, W. R.; Ando, K.; Powers, D. B.; Palkowitz, A. D.; Halterman, R. L. J. Am. Chem. Soc. 1990, 112, 6339. (c) Denmark, S. E.; Fu, J.; Lawler, M. J. J. Org. Chem. 2006, 71, 1523. (d) Kim, H.; Ho, S.; Leighton, J. L. J. Am. Chem. Soc. 2011, 133, 6517. (e) Meng, F.; Jang, H.; Jung, B.; Hoveyda, A. H. Angew. Chem., Int. Ed. 2013, 52, 5046.
- (18) For a review of N-methyl amino acids in peptide coupling, see: Humphrey, J. M.; Chamberlin, A. R. Chem. Rev. 1997, 97, 2243.
- (19) Devos, A.; Remion, J.; Frisque-Hesbain, A.-M.; Colens, A.; Ghosez, L. J. Chem. Soc., Chem. Commun. 1979, 1180.
- (20) For a review of acyl chlorides in peptide coupling, see: Prabhu, G.; Basavaprabhu; Narendra, N.; Vishwanatha, T. M.; Sureshbabu, V. V. Tetrahedron 2015, 71, 2785.
- (21) For selected examples of diene—ene RCM, see: (a) Burns, A. R.; McAllister, G. D.; Shanahan, S. E.; Taylor, R. J. K. Angew. Chem., Int. Ed. 2010, 49, 5574. (b) Biswas, K.; Lin, H.; Njardarson, J. T.; Chappell, M. D.; Chou, T.-C.; Guan, Y.; Tong, W. P.; He, L.; Horwitz, S. B.; Danishefsky, S. J. J. Am. Chem. Soc. 2002, 124, 9825. (c) Yang, Z.-Q.; Geng, X.; Solit, D.; Pratilas, C. A.; Rosen, N.; Danishefsky, S. J. J. Am. Chem. Soc. 2004, 126, 7881. (d) Lu, K.; Huang, M.; Xiang, Z.; Liu, Y.; Chen, J.; Yang, Z. Org. Lett. 2006, 8, 1193. (e) Wang, L.; Gong, J.; Deng, L.; Xiang, Z.; Chen, Z.; Wang, Y.; Chen, J.; Yang, Z. Org. Lett. 2009, 11, 1809. (f) Barluenga, S.; Lopez, P.; Moulin, E.; Winssinger, N. Angew. Chem., Int. Ed. 2004, 43, 3467.
- (22) For facile dichlorination of tyrosine esters, see: Hayashi, K.; Anzai, N.; Okayasu, I.; Endo, H. Immunosuppressant Containing *O*-(5-Amino-2-phenylbenzoxazol-7-yl)methyl-3,5-dichloro-L-tyrosine. Japan Patent JP 5470665 B1, April 16, 2014. In our modified procedure, complete protonation of the amino group by HCl is necessary to avoid side reactions. See the Supporting Information.